

# Gel/PCR DNA Fragments Extraction Kit

*For research use only*

**Sample:** up to 300 mg of agarose gel, up to 100 µl of PCR products

**Fragment Size:** 70 bp – 20 kb

**Recovery:** up to 95%

**Format:** spin column

**Operation Time:** 20 minutes (gel extraction), 10 minutes (PCR cleanup)

**Elution Volume:** 20-50 µl

**Storage:** dry at room temperature (15-25°C) for at least 1 year without showing any reduction in performance

**Geneaid**



CERTIFICATE NO. QAIC/TW/50077  
**ISO 9001:2008 QMS**

## Introduction

The Gel/PCR DNA Fragments Extraction Kit was designed to recover or concentrate DNA fragments (70 bp-20 kb) from agarose gel, PCR, or other enzymatic reactions in one convenient product. Chaotropic salt is used to dissolve agarose gel and denature enzymes. DNA fragments in chaotropic salt are bound by the glass fiber matrix of the spin column (1). Contaminants are removed with a Wash Buffer (containing ethanol) and the purified DNA fragments are eluted by a low salt Elution Buffer or TE. Salts, enzymes and unincorporated nucleotides can be effectively removed from the reaction mixture, without phenol extraction or alcohol precipitation. Typically, recoveries are up to 90% for Gel Extraction and up to 95% for PCR Clean Up. The eluted DNA is ready for use in PCR, Fluorescent or Radioactive Sequencing, Restriction Enzyme Digestion, DNA Labeling and Ligation.

## Quality Control

The quality of the Gel/PCR DNA Fragments Extraction Kit is tested on a lot-to-lot basis by isolating DNA fragments of various sizes from either aqueous solutions or agarose gel. The purified DNA is analyzed by electrophoresis.

## Kit Contents

Component	DF004	DF100	DF300
DF Buffer	3 ml	80 ml	240 ml
W1 Buffer	2 ml	45 ml	130 ml
Wash Buffer <sup>1</sup> (Add Ethanol)	1 ml (4 ml)	25 ml (100 ml)	50 ml + 25 ml (200 ml) (100 ml)
Elution Buffer	1 ml	6 ml	30 ml
DF Columns	4	100	300
2 ml Collection Tubes	4	100	300

<sup>1</sup>Add absolute ethanol (see the bottle label for volume) to Wash Buffer prior to initial use.

## Order Information

Post Reaction DNA Purification		
Product	Package Size	Cat. Number
GenepHlow™ Gel Extraction Kit	100/300 preps	DFG100/300
GenepHlow™ PCR Cleanup Kit	100/300 preps	DFC100/300
GenepHlow™ Gel/PCR Kit	100/300 preps	DFH100/300
GenepHlow™ DNA Cleanup Maxi Kit	10/25 preps	DFM010/025
Small DNA Fragments Extraction Kit	100/300 preps	DF101/301
Presto™ Max Gel/PCR Kit (Large DNA Fragments)	100/300 preps	DFL100/300
Presto™ 96 Well PCR Cleanup Kit	4/10 x 96 preps	96DFH04/10
Presto™ PCR Cleanup Kit 96 Well Binding Plate	10 plates	96DBP01
96-Well G-50 Gel Filtration Plate	4/10 x 96 rxns	CGP04/10
Gel Extraction Tool	25 pcs	GXT025

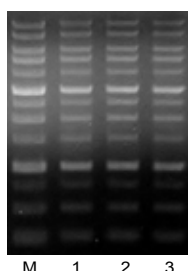
## Caution

DF Buffer contains guanidine thiocyanate. During operation, always wear a lab coat, disposable gloves, and protective goggles.

## References

(1) Vogelstein, B., and Gillespie, D. (1979) Proc. Natl. Acad. Sci. USA 76, 615

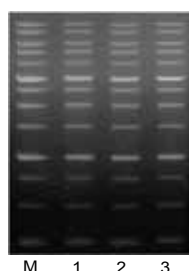
## Gel/PCR DNA Fragments Extraction Kit Functional Test Data



**Figure 1.** Gel slice DNA fragments ranging from 250 bp-10 kb were extracted using the Gel/PCR DNA Fragments Extraction Kit (total 1.0 µg, 3 replicates). The purified DNA from a 50 µl eluate was analyzed by electrophoresis on a 0.8% agarose gel.

M (Control) = Geneaid 1 Kb DNA Ladder

Test	DNA Conc.	260/280	Total DNA	Recovery
Control	---	---	1000 ng	---
Test-1	17.5 ng/µl	1.83	875 ng	87.5%
Test-2	16.4 ng/µl	1.84	820 ng	82%
Test-3	17.2 ng/µl	1.83	860 ng	86%



**Figure 2.** PCR product DNA fragments ranging from 250 bp-10 kb were extracted using the Gel/PCR DNA Fragments Extraction Kit (total 1.1 µg, 3 replicates). The purified DNA from a 50 µl eluate was confirmed by spectrophotometer and analyzed by electrophoresis on a 1% agarose gel.

M (Control) = Geneaid 1 Kb DNA Ladder

Test	DNA Conc.	260/280	Total DNA	Recovery
Control	---	---	1100 ng	---
Test-1	20.8 ng/µl	1.85	1040 ng	95%
Test-2	20.3 ng/µl	1.84	1015 ng	92%
Test-3	19.6 ng/µl	1.84	980 ng	89%

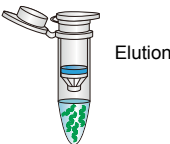
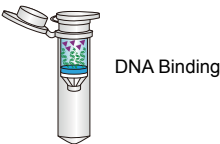
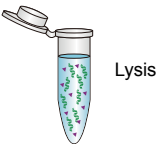
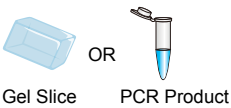
# Gel Extraction Protocol

## IMPORTANT BEFORE USE!

- Add absolute ethanol (see the bottle label for volume) to Wash Buffer prior to initial use
- Additional Requirements: absolute ethanol, microcentrifuge tubes

Gel Dissociation	<ul style="list-style-type: none"><li>• Excise the agarose gel slice containing relevant DNA fragments.</li><li>• Remove any extra agarose to minimize the size of the gel slice</li></ul> <p>NOTE: Using TAE buffer for gel formation is recommended for optimal DNA recovery.</p> <ul style="list-style-type: none"><li>• Transfer <b>up to 300 mg of the gel slice</b> to a 1.5 ml microcentrifuge tube.</li><li>• Add <b>500 µl of DF Buffer</b> to the sample and mix by vortex.</li><li>• Incubate at 55-60°C for 10-15 minutes to ensure the gel slice has been completely dissolved.</li></ul> <p>NOTE: During incubation, invert the tube every 2-3 minutes.</p> <ul style="list-style-type: none"><li>• Cool the dissolved sample mixture to room temperature.</li></ul>
Step 1 DNA Binding	<ul style="list-style-type: none"><li>• Place the <b>DF Column</b> in a <b>2 ml Collection Tube</b>.</li><li>• Transfer <b>800 µl of the sample mixture</b> to the <b>DF Column</b>.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds.</li><li>• Discard the flow-through and place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li></ul> <p>NOTE: If the sample mixture is more than 800 µl, repeat the DNA Binding step.</p>
Step 2 Wash	<ul style="list-style-type: none"><li>• Add <b>400 µl of W1 Buffer</b> into the <b>DF Column</b>.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds then discard the flow-through.</li><li>• Place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li><li>• Add <b>600 µl of Wash Buffer (make sure ethanol was added)</b> into the <b>DF Column</b>.</li><li>• Let stand for 1 minute at room temperature.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds then discard the flow-through.</li><li>• Place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li><li>• Centrifuge at 14-16,000 x g for 3 minutes to dry the column matrix.</li></ul>
Step 3 DNA Elution	<ul style="list-style-type: none"><li>• Transfer the dried <b>DF Column</b> to a new 1.5 ml microcentrifuge tube.</li><li>• Add <b>20-50 µl of Elution Buffer</b> or TE into the <b>CENTER</b> of the column matrix.</li><li>• Let stand for at least 2 minutes to ensure the <b>Elution Buffer</b> is completely absorbed.</li><li>• Centrifuge for 2 minutes at 14-16,000 x g to elute the purified DNA.</li></ul> <p>NOTE: Using pre-heated Elution Buffer (60°C) is recommended for eluting DNA fragments &gt;8kb.</p>

### Quick Protocol



# Gel Extraction For Sequencing Protocol

## IMPORTANT BEFORE USE!

- Add absolute ethanol (see the bottle label for volume) to Wash Buffer prior to initial use
- Additional Requirements: absolute ethanol, microcentrifuge tubes
- We recommend using the Gel Extraction For Sequencing Protocol as Guanidinium Chloride, a component of W1 Buffer, may interfere with sequencing reactions

Gel Dissociation	<ul style="list-style-type: none"><li>• Excise the agarose gel slice containing relevant DNA fragments.</li><li>• Remove any extra agarose to minimize the size of the gel slice</li></ul> <p>NOTE: Using TAE buffer for gel formation is recommended for optimal DNA recovery.</p> <ul style="list-style-type: none"><li>• Transfer up to 300 mg of the gel slice to a 1.5 ml microcentrifuge tube.</li><li>• Add <b>500 µl of DF Buffer</b> to the sample and mix by vortex.</li><li>• Incubate at 55-60°C for 10-15 minutes to ensure the gel slice has been completely dissolved.</li></ul> <p>NOTE: During incubation, invert the tube every 2-3 minutes.</p> <ul style="list-style-type: none"><li>• Cool the dissolved sample mixture to room temperature.</li></ul>
Step 1 DNA Binding	<ul style="list-style-type: none"><li>• Place the <b>DF Column</b> in a <b>2 ml Collection Tube</b>.</li><li>• Transfer <b>800 µl of the sample mixture</b> to the <b>DF Column</b>.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds.</li><li>• Discard the flow-through and place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li></ul> <p>NOTE: If the sample mixture is more than 800 µl, repeat the DNA Binding Step.</p>
Step 2 Wash	<ul style="list-style-type: none"><li>• Add <b>600 µl of Wash Buffer (make sure ethanol was added)</b> into the <b>DF Column</b> and let stand for 1 minute.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds then discard the flow-through.</li><li>• Place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li><li>• Add <b>600 µl of Wash Buffer (make sure ethanol was added)</b> into the <b>DF Column</b> and let stand for 1 minute.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds then discard the flow-through.</li><li>• Place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li><li>• Centrifuge at 14-16,000 x g for 3 minutes to dry the column matrix.</li></ul>
Step 3 DNA Elution	<ul style="list-style-type: none"><li>• Transfer the dried <b>DF Column</b> to a new 1.5 ml microcentrifuge tube.</li><li>• Add <b>20-50 µl of Elution Buffer</b> or TE into the <b>CENTER</b> of the column matrix.</li><li>• Let stand for at least 2 minutes to ensure the <b>Elution Buffer</b> is completely absorbed.</li><li>• Centrifuge for 2 minutes at 14-16,000 x g to elute the purified DNA.</li></ul> <p>NOTE: Using pre-heated Elution Buffer (60°C) is recommended for eluting DNA fragments &gt;8kb.</p>

# PCR Clean Up Protocol

## IMPORTANT BEFORE USE!

- Add absolute ethanol (see the bottle label for volume) to Wash Buffer prior to initial use
- Additional Requirements: absolute ethanol, microcentrifuge tubes

Sample Prep.	<ul style="list-style-type: none"><li>• Transfer <b>up to 100 µl of reaction product</b> to a 1.5 microcentrifuge tube.</li><li>• Add <b>5 volumes of DF Buffer</b> to 1 volume of the sample and mix by vortex.</li></ul>
Step 1 DNA Binding	<ul style="list-style-type: none"><li>• Place a <b>DF Column</b> in a <b>2 ml Collection Tube</b>.</li><li>• Transfer <b>the sample mixture</b> to the <b>DF Column</b>.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds.</li><li>• Discard the flow-through then place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li></ul>
Step 2 Wash	<ul style="list-style-type: none"><li>• Add <b>600 µl of Wash Buffer (make sure ethanol was added)</b> into the CENTER of the <b>DF Column</b>.</li><li>• Let stand for 1 minute at room temperature.</li><li>• Centrifuge at 14-16,000 x g for 30 seconds.</li><li>• Discard the flow-through and place the <b>DF Column</b> back in the <b>2 ml Collection Tube</b>.</li><li>• Centrifuge for 3 minutes at 14-16,000 x g to dry the column matrix.</li></ul>
Step 3 DNA Elution	<ul style="list-style-type: none"><li>• Transfer the dried <b>DF Column</b> to a new 1.5 ml microcentrifuge tube.</li><li>• Add <b>20-50 µl of Elution Buffer</b> or TE into the CENTER of the column matrix.</li><li>• Let stand for at least 2 minutes to ensure the <b>Elution Buffer</b> is completely absorbed.</li><li>• Centrifuge for 2 minutes at 14-16,000 x g to elute the purified DNA.</li></ul> <p>NOTE: Using pre-heated Elution Buffer (60°C) is recommended for eluting DNA fragments &gt;8kb.</p>

## Troubleshooting

Problem	Possible Reasons/Solution
Low Yield	<b>Gel slice did not dissolve completely</b> <ul style="list-style-type: none"><li>• The Gel slice was too big. If using more than 300 mg of gel slice, separate it into multiple tubes.</li><li>• Raise the incubation temperature to 60°C and extend the incubation time.</li></ul>
	<b>Incorrect DNA Elution Step</b> <ul style="list-style-type: none"><li>• Ensure that the Elution Buffer is completely absorbed after being added to the center of the DF Column.</li></ul>
	<b>Incomplete DNA Elution</b> <ul style="list-style-type: none"><li>• If the DNA fragments are larger than 8 kb, use pre-heated Elution Buffer (60-70°C) in the Elution Step to improve the elution efficiency.</li></ul>
Eluted DNA doesn't perform well in downstream applications	<b>Residual ethanol contamination</b> <ul style="list-style-type: none"><li>• Following the Wash Step, dry the DF Column with additional centrifugation at 14-16,000 x g for 5 minutes or incubate at 60°C for 5 minutes.</li></ul> <b>DNA was denatured (a smaller band appeared on gel analysis)</b> <ul style="list-style-type: none"><li>• Incubate the eluted DNA at 95°C for 2 minutes, and then cool down slowly to re-anneal the denatured DNA.</li></ul>
Low A260/A230	<ul style="list-style-type: none"><li>• In the wash step, repeat the 600 µl of Wash Buffer addition and let stand for 1 minute.</li></ul>

# Related DNA Extraction Products

Plasmid DNA Purification		
Product	Package Size	Catalogue Number
Presto™ Mini Plasmid Kit	100/300 preps	PDH100/300
Presto™ Midi Plasmid Kit	25 preps	PIF025
Presto™ Midi Plasmid Kit (Endotoxin Free)	25 preps	PIFE25
High-Speed Plasmid Mini Kit (10-50 Kb)	100/300 preps	PDL100/300
High-Speed Plasmid Advance Kit (50-100 ml)	25 preps	PA025
Geneaid™ Plasmid Mini Kit	40/100 preps	PAE040/100
Geneaid™ Plasmid Midi Kit	25 preps	PI025
Geneaid™ Plasmid Midi Kit (Endotoxin Free)	25 preps	PIE25
Presto™ Plasmid DNA Concentration Kit	250/500/1000 preps	PC0250/500/1000
Geneaid™ Plasmid Maxi Kit	10/25 preps	PM010/25
Geneaid™ Plasmid Maxi Kit (Endotoxin Free)	10/25 preps	PME10/25
Presto™ 96 Well Plasmid Kit	4/10 x 96 preps	96PDV04/10, 96PDC04/10
Presto™ Plasmid 96 Well Binding Plate	10 plates	96PBP01
Presto™ Plasmid 96 Well Filter Plate	10 plates	96PPF01
Post Reaction DNA Purification		
Product	Package Size	Catalogue Number
GenepHlow™ Gel Extraction Kit	100/300 preps	DFG100/300
GenepHlow™ PCR Cleanup Kit	100/300 preps	DFC100/300
GenepHlow™ Gel/PCR Kit	100/300 preps	DFH100/300
GenepHlow™ DNA Cleanup Maxi Kit	10/25 preps	DFM010/025
Small DNA Fragments Extraction Kit	100/300 preps	DF101/301
Presto™ Max Gel/PCR Kit (Large DNA Fragments)	100/300 preps	DFL100/300
Presto™ 96 Well PCR Cleanup Kit	4/10 x 96 preps	96DFH04/10
Presto™ PCR Cleanup Kit 96 Well Binding Plate	10 plates	96DBP01
DNA Pure Kit	100/300 preps	DP100/300
G-25 Gel Filtration Desalting Column	50 rxns	CG025
G-50 Gel Filtration Dye Terminator Removal Column	50 rxns	CG050
96-Well G-50 Gel Filtration Plate	4/10 x 96 rxns	CGP04/10
Gel Extraction Tool	25 pcs	GXT025
Genomic DNA Extraction and Purification		
Product	Package Size	Catalogue Number
Genomic DNA Mini Kit (Blood/Cultured Cell)	100/300 preps	GB100/300
Genomic DNA Midi Kit (Blood/Cultured Cell)	25 preps	GDI25
Genomic DNA Maxi Kit (Blood/Cultured Cell)	10/25 preps	GDM10/25
Genomic DNA Mini Kit (Tissue)	50/100/300 preps	GT050/100/300
gSYNC™ DNA Extraction Kit	100/300 preps	GS100/300
Genomic DNA Mini Kit (Plant)	100 preps	GP100
Genomic DNA Maxi Kit (Plant)	10/25 preps	GPM10/25
Geneaid™ DNA Isolation Kit (Blood)	100/1,000 rxns	GEB100/01K(+)
Geneaid™ DNA Isolation Kit (Bacteria)	300/3,000 rxns	GEE300/03K(+)
Geneaid™ DNA Isolation Kit (Tissue)	150/1,500 rxns	GET150/1.5K(+)
Geneaid™ DNA Isolation Kit (Cultured Cell)	150/1,500 rxns	GEC150/1.5K(+)
GENEzol™ DNA Reagent Plant	100/200 rxns	GR100/200
Presto™ Mini gDNA Yeast Kit	100/300 preps	GBY100/300
Presto™ Mini gDNA Bacteria Kit	100/300 preps	GBB100/101/300/301
Geneius™ Micro DNA Extraction Kit	100/300 preps	GMB100/300
Presto™ Buccal Swab gDNA Extraction Kit	100/300 preps	GSK100/300
DNA RNA Purification		
Product	Package Size	Catalogue Number
Presto™ DNA RNA Extraction Kit	50/100 preps	DR050/100

For additional product information please visit [www.geneaid.com](http://www.geneaid.com). Thank you!

